

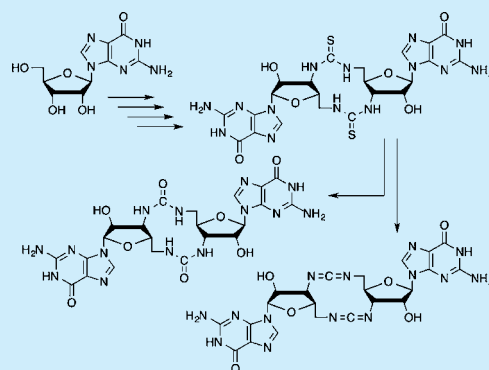
## Synthesis of c-di-GMP Analogs with Thiourea, Urea, Carbodiimide, and Guanidinium Linkages

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## Supporting Information

**ABSTRACT:** The first syntheses of neutral thiourea, urea, and carbodiimide analogs, along with two guanidinium analogs, of the bacterial signaling molecule cyclic diguanosine monophosphate (c-di-GMP) are reported. The key intermediate, obtained in nine steps, is a 3'-amino-5'-azido-3',5'-dideoxy derivative. The 5'-azide serves as a masked amine from which the amine is obtained by Staudinger reduction, while the 3'-amine is converted to an isothiocyanate that, while stable to chromatography, and Staudinger conditions, nevertheless reacts well with the 5'-amine.



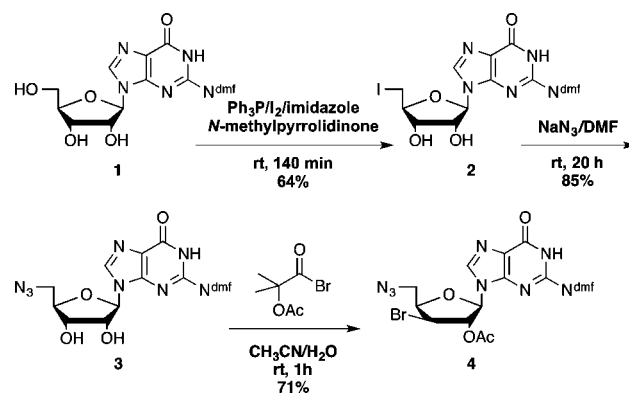
The bacterial signaling molecule cyclic diguanosine monophosphate (c-di-GMP) is responsible for regulating bacterial responses to a variety of environmental factors, including aggregation into the biofilm state.<sup>1–6</sup> Binding of c-di-GMP as a monomer and as a self-intercalated dimer to the PilZ domain proteins has been demonstrated.<sup>1,2,7,8</sup>

Activation of two different classes of riboswitches in noncoding regulatory mRNA domains also has been identified upon binding c-di-GMP.<sup>9–12</sup> Finally, c-di-GMP, among other cyclic dinucleotides, plays a role in triggering an innate immune response<sup>13,14</sup> through a transmembrane protein named STING in the innate immune sensing pathway, where a specific receptor for cyclic dinucleotides has been identified.<sup>15</sup>

A number of synthetic routes to c-di-GMP and its thiophosphate analogs have been reported.<sup>16–19</sup> Two analogs with a nonphosphate backbone have been prepared, one a methylphosphonate,<sup>20</sup> the other a carbamate,<sup>21</sup> but each lacks a 2'-hydroxyl group. An analog with a 2'-fluoro in place of the 2'-hydroxyl, with a phosphate backbone, was reported most recently.<sup>22</sup> The goal of the work reported below was to prepare c-di-GMP analogs with urea or urea related backbone linkages that should be stable to the bacterial phosphodiesterases that regulate c-di-GMP. The syntheses start with the introduction of nitrogen atoms to the guanosine 3' and 5' positions. The first steps are to prepare the 5'-azido-5'-deoxy derivative 3, as shown in Scheme 1.

The *N*<sup>2</sup>-dimethylformamidinium (dmf) derivative of guanosine, 1, was prepared by standard methods as described in detail in the Supporting Information. Preparation of 2 and 3 followed procedures reported for guanosine by Martin<sup>23</sup> and by Dean,<sup>24</sup> respectively. The major differences in this case were that heating was not required for the reaction of 2 with sodium

Scheme 1. Synthesis of Key Intermediate 4



azide and that 3 was readily isolated simply by the addition of methanol to the reaction mixture. The *N*<sup>2</sup>-dmf group was used in this synthesis, as it has been shown to be essential for the reaction of guanosine with  $\alpha$ -acetoxyisobutryl bromide.<sup>25</sup> The reaction of 3 with this reagent proceeded analogously to that reported for 1, with no degradation of the azido group under the acidic reaction conditions. In addition to the desired product, 4, a small amount of the 2'-Br isomer was produced, in the ratio of 92:8, by LC-MS. These isomers were not separable by silica chromatography, but 4 was readily crystallized from methylene chloride, which efficiently removed the 2'-Br isomer. No chromatography was required for the preparation of compounds 1–4, so that these reactions were conveniently

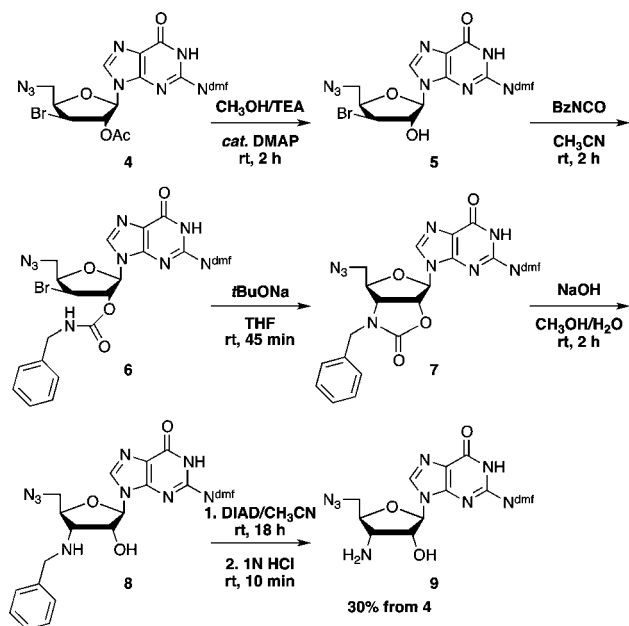
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carried out starting with 20 g of guanosine to give **4** in an overall yield of 38%.

The conversion of **4** to the 3'-amino-5'-azido derivative **9**, shown in Scheme 2, proceeded analogously to the preparation

**Scheme 2. Synthesis of 3'-Amino-5'-azido Derivative 9**



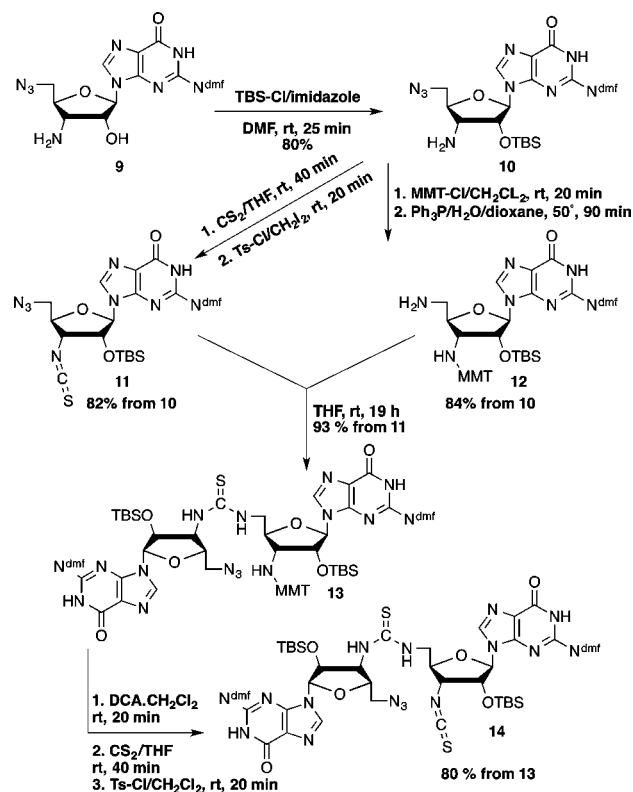
of 3'-amino-3'-deoxyguanosine reported by Zhang, although by using extensively altered conditions, and some different reagents, the reaction times were significantly reduced.<sup>26</sup> Catalytic DMAP in methanol with a few equivalents of TEA effected clean removal of the acetyl group from **4**. The reaction of **5** with benzylisocyanate in acetonitrile then proceeded in 2 h to give **6**. After investigating numerous reagents for cyclization to **7**, *t*BuONa in THF was found to give complete conversion in 45 min. Saponification of **7** to **8** by addition of 10 N NaOH to a methanol solution of **7** proceeded in 2 h. It is somewhat surprising that the *N*<sup>2</sup>-dmf group survived these strongly basic conditions with only minimal loss. After neutralization of the reaction mixture, **8** was isolated by extraction. The steps from **4** to **8** were carried out in one flask, without isolation of intermediates, and **8** did not need purification before conversion to **9**.

Because of the 5'-azide it was not possible to use reduction to debenzylate the 3'-amino group in **8**, and instead oxidation using diisopropylazodicarboxylate (DIAD) was employed.<sup>27</sup> This is a slow reaction that required overnight to give the corresponding imine (not shown). Hydrolysis to **9** was effected using 1 N HCl, within 10 min, again with minimal loss of the *N*<sup>2</sup>-dmf group. After neutralization of the reaction mixture with NaHCO<sub>3</sub>, **9** was isolated by extraction, in this case remaining in the aqueous phase, while excess reagent was removed in the organic phase. The purification of **9** was carried out by reversed phase chromatography using 10 mM aqueous ammonium bicarbonate and acetonitrile, to give **9** in a yield of 30% from **4**. Although the *N*<sup>2</sup>-dmf group survives limited time treatment with NaOH or HCl, it is slowly hydrolyzed by the ammonium bicarbonate eluant, so that solutions of **9** should not be allowed to stand for long periods of time after purification.

The derivatization of **9** for synthesis of the cyclic dimers required protection of the 2'-hydroxyl, conveniently done by

reaction with *tert*-butyldimethylsilyl chloride, as shown in Scheme 3. Addition of the TBS group makes **10** again

**Scheme 3. Synthesis of the Linear Dimer 14**



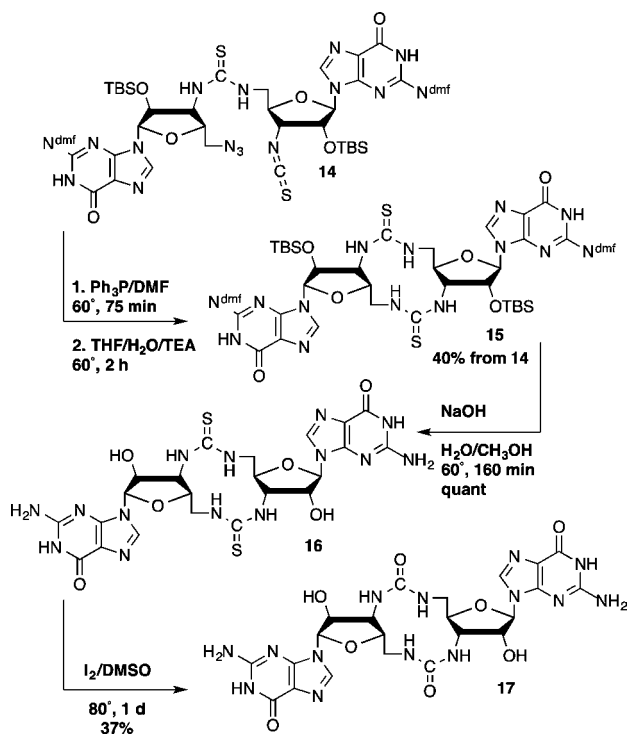
amenable to silica chromatography, and all of the subsequent intermediates were purified on silica using gradients of methanol (with 0.5% TEA for those with a free amino group or the acid labile monomethoxytrityl group) and methylene chloride. The strategy for synthesis of the linear and cyclic dimers was to elaborate the 3'-amino group into an isothiocyanate and to couple this to a 5'-amino group obtained by Staudinger reduction of the 5'-azide. The 3'-isothiocyanate is stable to silica chromatography, so that intermediates **11** and **14** are easily handled, but it does react well with the 5'-amino group. Thus the 5'-azide functions as a stable masked amino group that can be converted to the amine without harming the 3'-isothiocyanate of **14**.

Formation of the 3'-isothiocyanate derivative **11** was carried out by reaction of **10** with carbon disulfide followed by reaction of the resulting dithiocarbamate (not shown) with tosyl chloride or benzenesulfonyl chloride.<sup>28</sup> This was done as a two-step procedure using a 10-fold excess of CS<sub>2</sub> in the first step that was readily removed on a rotary evaporator before reaction with the sulfonyl chloride. The 5'-amino nucleoside **12** was obtained by Staudinger reduction after protection of the 3'-amino group of **10** by reaction with monomethoxytrityl chloride. Condensation of **11** and **12** in THF at room temperature gave clean conversion to the linear dimer **13** within 17 h in 93% yield. The monomethoxytrityl group was removed using dichloroacetic acid (DCA), and the amino group converted to an isothiocyanate to give **14** by the same two-step procedure used for preparation of **11**.

Cyclization of the linear dimer **14** to the cyclic dimer **15** was effected by a two-step sequence starting with reaction of **14**

with triphenyl phosphine to give the azine (not shown, but sufficiently stable to be clearly visible by LC-MS), followed by dilution of the reaction mixture with THF/water/TEA and heating at 60 °C for 2 h (Scheme 4). Although LC-MS showed

Scheme 4. Syntheses of Thiourea 16 and Urea 17

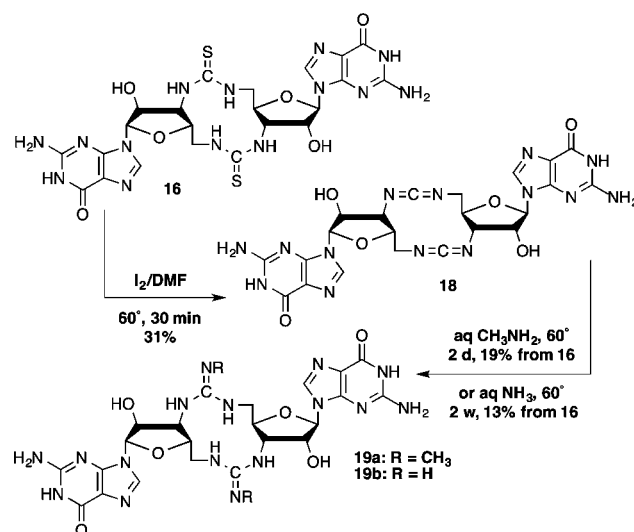


that the protected cyclic dimer **15** was the only significant product, there were a number of small impurities visible, possibly oligomers from an intermolecular reaction, even under the dilute conditions of the cyclization. It was again possible to purify **15** by silica chromatography, using a steep gradient of methanol in methylene chloride. The isolated yield for **15** was only 40%, presumably because of competing intermolecular reactions or degradation.

The deprotection of **15** to the cyclic thiourea **16** was effected using 2 N NaOH in methanol/water (1:1). Under these conditions the TBS groups were removed in minutes, at room temperature, while the *N*<sup>2</sup>-dmf groups required heating at 60 °C for 2 h to effect removal, consistent with the surprising stability noted earlier. Neutralization of the reaction mixture with either 1 N HCl or acetic acid caused precipitation of **16**, which was isolated by filtration in quantitative yield. Of the many potential routes for conversion of thioureas to ureas,<sup>29</sup> reaction of **16** with DMSO and catalytic iodine, at 80 °C, was employed.<sup>30</sup> This is a simple, if slow, procedure that does not involve metals or unusual conditions and gave clean conversion to **17**, in 37% yield.

The reaction of **16** with iodine, this time in DMF at room temperature with triethylamine, was also effective for preparation of the carbodiimide **18** (Scheme 5).<sup>31</sup> Although this reaction is reported to require aryl thioureas,<sup>31</sup> it worked well for preparation of **18**. The reaction of carbodiimides with amines for synthesis of guanidines is well-known,<sup>32</sup> and aqueous methylamine and aqueous ammonia gave **19a** and **19b**, respectively, although slowly and in modest yields.

Scheme 5. Syntheses of Carbodiimide 18 and Guanidines 19a and 19b



The carbodiimide **18** proved to be sufficiently stable to be handled and purified using the same conditions used for **17** and **19a/b**. All of these compounds have poor solubility in water, but are soluble in 0.1 N NaOH. Purification of each was done by RP chromatography using 0.1 N NaOH and methanol. Neutralization of the product fractions using CO<sub>2</sub> gas gave each compound as a white solid easily isolated by filtration. The preparations of **17**, **18**, and **19a/b** were carried out on small scales only and were not optimized.

Recent reports of activation of the innate immune system with the 2'/3' isomers of cyclic GMP-AMP (cGAMP)<sup>33–35</sup> provide a new impetus for preparation of cyclic dinucleotides (CDNs) and their analogs. The compounds reported here are the first examples of a new class of CDN analogs that possess a urea, or urea related, backbone.

## ■ ASSOCIATED CONTENT

### Supporting Information

Synthetic procedures, HPLC, <sup>1</sup>H, <sup>13</sup>C NMR, and UV spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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### Notes

The authors declare no competing financial interest.

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## ■ REFERENCES

- (1) Hengge, R. *Nat. Rev. Microbiol.* **2009**, *7*, 263–273.
- (2) Krasteva, P. V.; Giglio, K. M.; Sondermann, H. *Protein Sci.* **2012**, *21*, 929–948.
- (3) Mills, E.; Pultz, I. S.; Kulasekara, H. D.; Miller, S. I. *Cell. Microbiol.* **2011**, *13*, 1122–1129.
- (4) Povolotsky, T. L.; Hengge, R. *J. Biotechnol.* **2012**, *160*, 10–16.

- (5) Quin, M. B.; Berrisford, J. M.; Newman, J. A.; Baslé, A.; Lewis, R. J.; Marles-Wright, J. *Structure* **2012**, *20*, 350–363.
- (6) Sondermann, H.; Shikuma, N. J.; Yildiz, F. H. *Curr. Opin. Microbiol.* **2012**, *15*, 140–146.
- (7) Schirmer, T.; Jenal, U. *Nat. Rev. Micro.* **2009**, *7*, 724–735.
- (8) Ko, J.; Ryu, K.-S.; Kim, H.; Shin, J.-S.; Lee, J.-O.; Cheong, C.; Choi, B.-S. *J. Mol. Biol.* **2010**, *398*, 97–110.
- (9) Shanahan, C. A.; Gaffney, B. L.; Jones, R. A.; Strobel, S. A. *J. Am. Chem. Soc.* **2011**, *133*, 15578–15592.
- (10) Smith, K. D.; Lipchock, S. V.; Ames, T. D.; Wang, J.; Breaker, R. R.; Strobel, S. A. *Nat. Struct. Mol. Biol.* **2009**, *16*, 1218–1223.
- (11) Smith, K. D.; Shanahan, C. A.; Moore, E. L.; Simon, A. C.; Strobel, S. A. *Proc. Natl. Acad. Sci. U.S.A.* **2011**, *108*, 7757–7762.
- (12) Sudarsan, N.; Lee, E. R.; Weinberg, Z.; Moy, R. H.; Kim, J. N.; Link, K. H.; Breaker, R. R. *Science (Wash.)* **2008**, *321*, 411–413.
- (13) Karaolis, D. K. R.; Means, T. K.; Yang, D.; Takahashi, M.; Yoshimura, T.; Muraille, E.; Philpott, D.; Schroeder, J. T.; Hyodo, M.; Hayakawa, Y.; Talbot, B. G.; Brouillette, E.; Malouin, F. *J. Immunol.* **2007**, *178*, 2171–2181.
- (14) Woodward, J. J.; Iavarone, A. T.; Portnoy, D. A. *Science* **2010**, *328*, 1703–1705.
- (15) Burdette, D. L.; Monroe, K. M.; Sotelo-Troha, K.; Iwig, J. S.; Eckert, B.; Hyodo, M.; Hayakawa, Y.; Vance, R. E. *Nature* **2011**, *478*, 515–518.
- (16) Gaffney, B. L.; Veliath, E.; Zhao, J.; Jones, R. A. *Org. Lett.* **2010**, *12*, 3269–3271.
- (17) Kiburu, I.; Shurer, A.; Yan, L.; Sintim, H. O. *Mol. Biosyst.* **2008**, *4*, 518–520.
- (18) Yan, H.; Wang, X.; KuoLee, R.; Chen, W. *Biorg. Med. Chem. Lett.* **2008**, *18*, 5631–5634.
- (19) Hyodo, M.; Sato, Y.; Hayakawa, Y. *Tetrahedron* **2006**, *62*, 3089–3094.
- (20) Shanahan, C. A.; Gaffney, B. L.; Jones, R. A.; Strobel, S. A. *Biochemistry* **2013**, *52*, 365–377.
- (21) Kline, T.; Jackson, S. R.; Deng, W.; Verlinde, C. L. M. J.; Miller, S. I. *Nucleosides Nucleotides Nucl. Acids* **2008**, *27*, 1282–1300.
- (22) Zhou, J.; Watt, S.; Wang, J.; Nakayama, S.; Syre, D. A.; Lam, Y.; Lee, V. T.; Sintim, H. O. *Biorg. Med. Chem.* **2013**, *21*, 4396–4404.
- (23) McGee, D. P. C.; Martin, J. C. *Can. J. Chem.* **1986**, *64*, 1885–1889.
- (24) Dean, D. K. *Synth. Commun.* **2002**, *32*, 1517–1521.
- (25) He, G.-X.; Bischofberger, N. *Tetrahedron Lett.* **1995**, *36*, 6991–6994.
- (26) Zhang, L.; Cui, Z.; Zhang, B. *Helv. Chim. Acta* **2003**, *86*, 703–710.
- (27) Kroutil, J.; Trnka, T.; Černý, M. *Synthesis* **2004**, 446–450.
- (28) Wong, R.; Dolman, S. J. *J. Org. Chem.* **2007**, *72*, 3639–3971.
- (29) Sahu, S.; Sahoo, P. R.; Patel, S.; Mishra, B. K. *J. Sulfur Chem.* **2011**, *32*, 171–197.
- (30) Mikołajczyk, M.; Łuczak, J. *Synthesis* **1975**, 114–115.
- (31) Ali, A. R.; Ghosh, H.; Patel, B. K. *Tetrahedron Lett.* **2010**, *51*, 1019–1021.
- (32) Katritzky, A. R.; Rogovoy, B. V. *ARKIVOC* **2005**, 2005, 49–87.
- (33) Gao, P.; Ascano, M.; Wu, Y.; Barchet, W.; Gaffney, B. L.; Zillinger, T.; Serganov, A. A.; Liu, Y.; Jones, R. A.; Hartmann, G.; Tuschl, T.; Patel, D. J. *Cell* **2013**, *153*, 1094–1107.
- (34) Wu, J.; Sun, L.; Chen, X.; Du, F.; Shi, H.; Chen, C.; Chen, Z. J. *Science* **2013**, *339*, 826–830.
- (35) Ablasser, A.; Goldeck, M.; Cavlar, T.; Deimling, T.; Witte, G.; Röhl, I.; Hopfner, K.-P.; Ludwig, J.; Hornung, V. *Nature* **2013**, *498*, 380–384.